

STRUCTURE OF RK-397, A NEW OXO  
 PENTAENE ANTIBIOTIC

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RK-397 is a new antibiotic produced by *Streptomyces* sp. 87-397. The antibiotic showed antitumor, antifungal and antibacterial activities. The studies of taxonomy, fermentation, isolation, physico-chemical properties and biological activities are described in the preceding paper<sup>1)</sup>. In this paper, we report the structural determination of RK-397.

RK-397 is a yellow powder with mp 157~163°C (dec) and has the molecular formula C<sub>35</sub>H<sub>56</sub>O<sub>10</sub> which was determined by HRFAB-MS (*m/z* 637.3930, MH<sup>+</sup>). The UV spectrum exhibited characteristic absorption of the oxo pentaene chromophore, λ<sub>max</sub> (in MeOH) 360 nm, and suggested that RK-397 belongs the oxo polyene macrolide group of antibiotics, such as mycotocins<sup>2)</sup>, roxaticin<sup>3)</sup>, roflamycoin<sup>4)</sup> and dermostatins<sup>5)</sup>. The <sup>1</sup>H NMR spectrum (Table 1) showed the ten olefinic protons of the oxo pentaene moiety, as well as two protons of an isolated olefin. The <sup>13</sup>C NMR and DEPT spectrum (Table 1) exhibited the signals of one carbonyl carbon, three methyl carbons, eight methylene carbons and twenty-three methine carbons which consist of twelve olefinic, nine oxygenated and two ordinary methine carbons. Two partial structures, C-1~C-12 and C-27~C-35, were assigned by <sup>1</sup>H-<sup>1</sup>H COSY and <sup>13</sup>C-<sup>1</sup>H COSY. The stereochemistry of the six double bonds was determined as all *E* from the large coupling constants (*J*=14.9 Hz for the pentaene moiety and *J*=15.7 Hz for H-28 and H-29). The remaining portion (C-13~C-26) contains seven methylenes (45~48 ppm) and seven oxygenated methines (65~69 ppm). These <sup>13</sup>C chemical shifts suggested that the presence of a skipped polyol for the C-13~C-26 portion<sup>6,7)</sup>. Careful analyses of <sup>1</sup>H-<sup>1</sup>H HOHAHA and HMBC<sup>8)</sup> spectra in addition to <sup>1</sup>H-<sup>1</sup>H COSY allowed full assignment of <sup>1</sup>H and <sup>13</sup>C NMR signals. Further, the 2D <sup>13</sup>C-<sup>1</sup>H HOHAHA spectrum was measured and this data

supported the assignment of the crowded proton signals for the skipped polyol portion (Table 2). In the HMBC spectrum summarized in Fig. 1, <sup>1</sup>H-<sup>13</sup>C long range couplings from H-2, H-3 and H-31 to the carbonyl carbon (C-1 at 169.07 ppm) were observed. The chemical shift of the methine proton H-31 (4.78 ppm) indicated that this methine is acyloxy methine. Consequently, the planar structure of RK-397 (1) was determined to be the 32-membered macrolide depicted in Fig. 2.

 Table 1. <sup>13</sup>C (150 MHz) and <sup>1</sup>H NMR (600 MHz) data for RK-397 (1)<sup>a,b</sup>.

No.	C	H <sup>c</sup>
1	169.07	—
2	121.40	5.89 (d, 14.9)
3	146.84	7.30 (dd, 14.9, 11.7)
4	131.07	6.44 (dd, 14.9, 11.7)
5	142.84	6.70 (dd, 14.9, 11.3)
6	132.86	6.37 (dd, 14.9, 11.3)
7	139.30	6.51 (dd, 14.9, 10.9)
8	132.38	6.29 (dd, 14.9, 10.9)
9	137.12	6.40 (dd, 14.9, 10.9)
10	133.79	6.22 (dd, 14.9, 10.1)
11	134.19	5.85 (ddd, 14.9, 10.1, 5.3)
12	43.95	2.63 (m), 2.24 (ddd, 13.7, 10.1, 10.0)
13	69.35	3.96 (m)
14	46.92	1.68~1.59 (m)
15	67.53	3.82 (m)
16	48.03	1.70 (ddd, 13.7, 8.2, 4.8), 1.38 (ddd, 13.7, 8.9, 4.4)
17	66.36	3.89 (m)
18	45.34	1.46 (ddd, 14.1, 9.6, 3.2), 1.20 (ddd, 14.1, 10.1, 2.4)
19	65.09	4.07 (m)
20	47.37	1.33 (m), 1.27 (m)
21	68.98	3.94 (m)
22	46.29	1.33 (m), 1.25 (m)
23	68.55	4.04 (m)
24	46.37	1.44 (ddd, 14.1, 10.1, 8.4), 1.33 (m)
25	67.75	3.91 (m)
26	46.78	1.50 (ddd, 14.1, 10.1, 8.4), 1.31 (m)
27	71.56	4.22 (m)
28	133.57	5.53 (ddd, 15.7, 4.8, 1.6)
29	132.35	5.64 (ddd, 15.7, 5.2, 1.2)
30	37.48	2.64 (m)
31	81.93	4.78 (dd, 10.0, 2.6)
32	30.67	1.92 (dq, 10.0, 6.9, 6.8)
33	19.15	0.98 (d, 6.9)
34	11.43	1.06 (d, 6.9)
35	20.24	0.88 (d, 6.8)

<sup>a</sup> Chemical shifts are given in ppm from TMS as internal standard.

<sup>b</sup> CD<sub>3</sub>OD as solvent.

<sup>c</sup> Multiplicity and coupling constants (*J* in Hz) are given in parentheses.

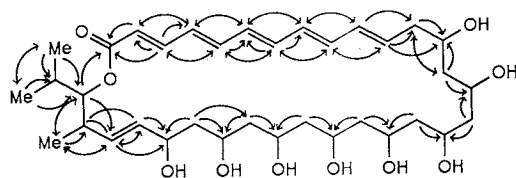
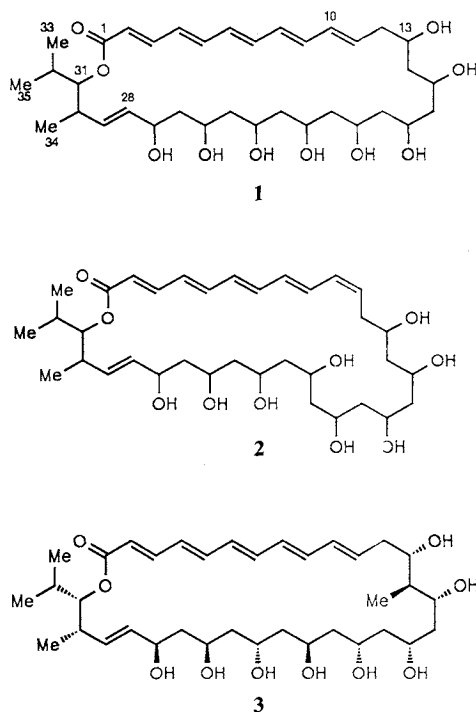
Table 2. Cross peaks observed in the 2D  $^{13}\text{C}$ - $^1\text{H}$  HOHAHA experiment for RK-397 (1)<sup>a,b</sup>.

C	H	C	H
C-4	3-H, 5-H	C-23	22-H, 24-H
C-5	6-H	C-27	26-H
C-7	6-H	C-28	29-H
C-9	10-H	C-30	34-H
C-10	9-H, 11-H	C-31	32-H, 33-H, 35-H
C-12	13-H	C-32	30-H, 31-H, 33-H,
C-14	15-H		35-H
C-15	14-H, 16-H	C-33	35-H
C-17	16-H, 18-H	C-34	30-H
C-19	18-H, 20-H	C-35	33-H

<sup>a</sup> The  $^{13}\text{C}$ - $^1\text{H}$  HOHAHA spectrum was recorded in the absorptive mode, using mixing time 60 mseconds, by 600 MHz spectroscopy.

<sup>b</sup> Cross peaks of direct coupling ( $^1J_{\text{CH}}$ ) were eliminated.

Fig. 1. Long range C-H coupling observed for RK-397 (1) in the HMBC experiment.

Fig. 2. Structures of RK-397 (1), 10,11-*cis* RK-397 (2) and mycoticin A (3).Table 3.  $^1\text{H}$  NMR data (400 MHz) for 10,11-*cis* RK-397 (2)<sup>a,b</sup>.

No.	H <sup>c,d</sup>
2	5.94 (d, 15.1)
3	7.33 (dd, 15.1, 11.7)
4	6.46 (dd, 15.1, 11.7)
5	6.75 (dd, 15.1, 11.2)
6	6.41 (dd, 14.7, 11.2)
7	6.60 (dd, 14.7, 11.2)
8	6.37 (dd, 14.7, 11.2)
9	6.70 (dd, 14.7, 11.7)
10	6.25 (dd, 11.7, 11.7)
11	5.72 (ddd, 11.7, 11.7, 3.9)
12	2.83 (ddd, 12.7, 11.7, 2.0), 2.33 (m)
13	4.14 (m)
27	4.20 (m)
28	5.52 (m)
29	5.55 (m)
30	2.61 (m)
31	4.76 (dd, 9.7, 1.4)
32	1.95 (m)
33	0.97 (d, 6.8)
34	1.05 (d, 6.9)
35	0.86 (d, 6.9)

<sup>a</sup> Chemical shifts are given in ppm from TMS as internal standard.

<sup>b</sup>  $\text{CD}_3\text{OD}$  as solvent.

<sup>c</sup> Multiplicity and coupling constants ( $J$  in Hz) are given in parentheses.

<sup>d</sup> Assignments of 14-H ~ 26-H were not determined. Signals of 15, 17, 19, 23-H, and 25-H were observed at 4.25 ~ 3.60 ppm and signals of 14, 16, 18, 20, 22, 24-H and 26-H were observed at 1.90 ~ 1.10 ppm.

RK-397 was labile to light and converted to some geometrical isomers. One of the isomers, compound **2**, was partially purified and analyzed by NMR spectroscopy. Compound **2** and RK-397 (**1**) have the same molecular formulas, which were determined by HRFAB-MS. The  $^1\text{H}$  NMR spectrum (Table 3) of **2** was similar to that of RK-397, but some difference was observed in the signals of the pentaene moiety. Sequential assignment of protons of **2** from the doublet signal of H-2 at 5.94 ppm was carried out by  $^1\text{H}$ - $^1\text{H}$  COSY. The vicinal coupling constant between H-10 and H-11 was  $J=11.7\text{ Hz}$ , and the stereochemistry of this double bond was assigned to *cis* configuration. Other double bonds were assigned to *trans* stereochemistry from large coupling constants. The structure of compound **2** was thus determined to be the 10,11-*cis* isomer of RK-397. Recently, fulongmycin<sup>9)</sup> which has the planar structure of 10,11-*cis* mycoticin was reported.

RK-397 (**1**) is a new member of the oxo polyene

macrolide antibiotics which include mycotocins A(3) and B<sup>2</sup>), roxaticin<sup>3</sup>), roflamycin<sup>4</sup>), dermostatins A and B<sup>5</sup>), fulongmycin<sup>9</sup>), flavofungins<sup>10</sup>), and surgumycin<sup>11</sup>). The configurations of the oxo polyene antibiotics, roxaticin<sup>3</sup>) and mycotocins<sup>12~14</sup>) were determined by X-ray analysis and synthesis, respectively. The planar structure of RK-397 corresponds to 14-demethyl-mycoticin A. The NMR data for RK-397 gave some information about the stereochemistry, but the determination of the stereochemistry for the polyol portion of RK-397 needs further experiments<sup>15</sup>). The NMR data for vicinal coupling constants (Table 1) indicated that the all *E* conjugated pentaene is extended *s-trans* conformation and the conformation of terminal portion (C-28~C-35) is the same as that of mycotocin A ( $J_{29,30} = 5.4$ ,  $J_{30,31} = 2.5$ ,  $J_{31,32} = 10.0$  Hz for 3<sup>15</sup>) and roxaticin<sup>3</sup>).

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